

[0306] Yen J Y et al., Therapeutics of Ebola hemorrhagic fever: whole-genome transcriptional analysis of successful disease mitigation. *J Infect Dis.* 2011 November; 204 Suppl 3:S1043-52.

[0307] Zabihollahi R et al., Inhibition of HIV and HSV infection by vaginal lactobacilli in vitro and in vivo. *Daru.* 2012 Oct. 15; 20(1):53.

[0308] Zhang W J et al., Associated changes in the transcription levels of IL-17A and tight junction-associated genes in the duodenal mucosa of rhesus macaques repeatedly exposed to simian/human immunodeficiency virus. *Exp Mol Pathol.* 2014 Jul. 14.

[0309] Zhou X I et al., A novel helper-dependent adenovirus-based cell culture model for Hepatitis C virus replication and production. *Virol J.* 2013 Aug. 30; 10:273. What is claimed is:

1. A method for treating or ameliorating a virus infection, comprising administering a composition that comprises a cultured placental adherent stromal cell (ASC), thereby treating or ameliorating a virus infection.

2. A method for treating, preventing, or ameliorating a complication of a virus infection, comprising administering a composition that comprises a cultured placental adherent stromal cell (ASC), thereby treating, preventing, or ameliorating a complication of a virus infection.

3. The method of claim 1, where said virus is selected from HIV-1, HCV, HBV, HSV-1, HSV-2, Dengue virus, Marburg virus, Ebola virus, yellow fever virus, Lassa virus, Crimean-Congo HFV, and Rift Valley virus.

4. The method of claim 1, where said composition is an injected composition.

5. The method of claim 1, wherein said placental ASC have been incubated on a 2D substrate.

6. The method of claim 1, wherein said placental ASC have been incubated on a 3D substrate.

7. The method of claim 6, wherein said placental ASC have been incubated on a 2D substrate, prior to incubating on a 3D substrate.

8. The method of claim 7, wherein said 3D culture substrate comprises a fibrous matrix, comprising a synthetic adherent material, where said synthetic adherent material is

selected from the group consisting of a polyester, a polypropylene, a polyalkylene, a polyfluorochloroethylene, a polyvinyl chloride, a polystyrene, and a polysulfone.

9. The method of claim 8, wherein said 3D culture apparatus is in form of microcarriers, wherein said microcarriers are disposed in a bioreactor.

10. The method of claim 1, wherein said placental ASC is allogeneic to said subject.

11. The method of claim 1, wherein the composition is intramuscularly injected.

12. The method of claim 1, comprising 100-600 million of said placental ASC, for an adult subject.

13. The method of claim 1, wherein said composition comprises:

- a. a first pharmaceutical composition, comprising allogeneic placental ASC from a first donor; and
- b. a second pharmaceutical composition, comprising allogeneic placental ASC from a second donor, wherein said second donor differs from said first donor in at least one allele group of human leukocyte antigen (HLA)-A or human leukocyte antigen (HLA)-B.

14. The method of claim 13, wherein said second pharmaceutical composition administered to said subject at least 7 days after said first pharmaceutical composition is administered.

15. The method of claim 1, wherein said ASC express a marker selected from the group consisting of CD73, CD90, CD29 and CD105.

16. The method of claim 1, wherein said ASC do not express a marker selected from the group consisting of CD3, CD4, CD11b, CD14, CD19, and CD34.

17. The method of claim 1, wherein said ASC do not express a marker selected from the group consisting of CD3, CD4, CD34, CD39, and CD106.

18. The method of claim 17, wherein less than 50% of said ASC express CD200.

19. The method of claim 17, wherein more than 50% of said ASC express CD200.

20. The method of claim 17, wherein more than 50% of said ASC express CD141.

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